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# CHLORIDE-BICARBONATE EXCHANGE IN THE URINARY BLADDER OF THE TURTLE

## INDEPENDENCE FROM SODIUM ION

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### Summary

The rates of  $\text{Cl}^-$  absorption and  $\text{HCO}_3^-$  secretion were not different in turtle urinary bladders bathed in  $\text{Na}^+$ -containing and  $\text{Na}^+$ -free solutions.

These results in turtle bladder are inconsistent with  $\text{Na}^+$ -anion cotransport but can be accounted for by a  $\text{Cl}^-/\text{HCO}_3^-$  exchange system.

Transport of  $\text{Cl}^-$  occurs by three mechanisms in epithelial tissues: passively down electrochemical gradients, by  $\text{Na}^+\text{-Cl}^-$  cotransport and by  $\text{Cl}^-/\text{HCO}_3^-$  exchange [1]. The urinary bladder of the turtle (*Pseudemys scripta*) transport  $\text{Cl}^-$  from the mucosal solution to the serosal solution [2, 3] and  $\text{HCO}_3^-$  from serosal→mucosal [3, 4]. Earlier studies have shown that the absorption of  $\text{Cl}^-$  is independent of  $\text{Na}^+$  [2]. In the presence of  $\text{Na}^+$ ,  $\text{Cl}^-$  absorption and  $\text{HCO}_3^-$  secretion occur by a coupled anion exchange system [3]. The present study was designed to determine whether the stoichiometry of the exchange process is altered in the absence of  $\text{Na}^+$ .

The experimental methods are described in detail in previous publications [3, 5]. In brief, hemibladders were mounted on lucite chambers and bathed on both surfaces by a  $\text{HCO}_3^-$ -free Ringer's solution containing 116 mM  $\text{Na}^+$  or a  $\text{HCO}_3^-$ -free Ringer's solution with the  $\text{Na}^+$  replaced by  $\text{Cs}^+$  [5].  $\text{HCO}_3^-$  secretion was measured by pH-stat titration and mucosal→serosal movement of  $\text{Cl}^-$  by flux of radioactive  $\text{Cl}^-$  ( $^{36}\text{Cl}$ ).

The mucosal pH was lowered until  $\text{H}^+$  secretion was abolished [6] and the rate of unidirectional mucosal→serosal  $\text{Cl}^-$  flux was determined. In the absence of exogenous  $\text{HCO}_3^-$  in the bathing solutions the mucosal→serosal  $\text{Cl}^-$  flux is equivalent to the serosal→mucosal  $\text{Cl}^-$  flux [7]. The addition of

$\text{HCO}_3^-$  to the serosal solution causes an increase in the mucosal→serosal  $\text{Cl}^-$  flux but has no effect on the serosal→mucosal flux [7]. Therefore, the mucosal→serosal  $\text{Cl}^-$  flux in the absence of  $\text{HCO}_3^-$  has been used as an estimate of passive flux and the increment in the mucosal→serosal flux of  $\text{Cl}^-$  after  $\text{HCO}_3^-$  addition is an estimate of the exchange flux.

After measurement of mucosal→serosal  $\text{Cl}^-$  flux in the absence of  $\text{HCO}_3^-$ , the serosal solution was replaced with a Ringer's solution containing 20 mM  $\text{HCO}_3^-$ .  $\text{HCO}_3^-$  secretion (serosal→mucosal) and mucosal→serosal  $\text{Cl}^-$  flux were determined following a 1 h equilibration period. At the completion of the flux measurements,  $10^{-4}$  M amiloride was added to the mucosal solution. Under such conditions (i.e. complete inhibition of electrogenic  $\text{Na}^+$  and  $\text{H}^+$  transport), the remaining current is unaffected by cyanide or deoxyglycose (Cohen, L.H. personal communication). This current is probably due to passive diffusion of ions down their chemical gradients. Chemical gradients exist only for  $\text{HCO}_3^-$  and  $\text{SO}_4^{2-}$  in the present experiments. The  $\text{SO}_4^{2-}$  gradient (10 mM) would account for less than 10% of the diffusion current based on data for  $\text{SO}_4^{2-}$  fluxes in turtle bladder [8]. Therefore, the post-amiloride current has been used as an estimate of the passive  $\text{HCO}_3^-$  flow.

The mucosal pH at which net acid secretion was abolished was  $4.58 \pm 0.10$  in  $\text{Na}^+$ -containing Ringer's and  $4.55 \pm 0.12$  in  $\text{Na}^+$ -free Ringer's. Table I shows that the total  $\text{HCO}_3^-$  movement from serosal→mucosal was similar in  $\text{Na}^+$ -containing and  $\text{Na}^+$ -free Ringer's. The total  $\text{Cl}^-$  flux from mucosal→serosal also was unaffected by the absence of  $\text{Na}^+$ . The total  $\text{HCO}_3^-$  movement was equal to the total  $\text{Cl}^-$  flux during both  $\text{Na}^+$  and  $\text{Na}^+$ -free conditions.

The total anion fluxes can be dissected into a passive component and an exchange component. Table II shows that the passive portion of the  $\text{Cl}^-$

TABLE I

EFFECT OF  $\text{Na}^+$  ON UNIDIRECTIONAL ANION MOVEMENTS

Mean values  $\pm$  S.E. of anion movements. 20 mM  $\text{HCO}_3^-$  and 1%  $\text{CO}_2$  were present in the serosal solution.

The area of exposed tissue was 8 cm<sup>2</sup>

	Bathing media	
	Na (n = 5)	Na-free (n = 7)
mucosal→serosal $\text{Cl}^-$ ( $\mu\text{mol/h}$ )	$0.53 \pm 0.12$	$0.56 \pm 0.08$
serosal→mucosal $\text{HCO}_3^-$ ( $\mu\text{mol/h}$ )	$0.58 \pm 0.08$	$0.55 \pm 0.05$

TABLE II

EFFECT OF  $\text{Na}^+$  ON PASSIVE ANION MOVEMENTS

Mean values  $\pm$  S.E. of passive anion movements. Passive  $\text{Cl}^-$  movement was estimated as the mucosal→serosal  $\text{Cl}^-$  flux in the absence of serosal  $\text{HCO}_3^-$ . Passive  $\text{HCO}_3^-$  movement was estimated as the current remaining after blockade of  $\text{Na}^+$  transport ( $10^{-4}$  M amiloride to the mucosal solution) and acidification (by lowering mucosal pH). Same tissues as in Table I.

	Bathing media	
	Na (n = 5)	Na-free (n = 7)
mucosal→serosal $\text{Cl}^-$ ( $\mu\text{mol/h}$ )	$0.30 \pm 0.06$	$0.28 \pm 0.04$
serosal→mucosal $\text{HCO}_3^-$ ( $\mu\text{mol/h}$ )	$0.33 \pm 0.09$	$0.23 \pm 0.04$

flux and the diffusional  $\text{HCO}_3^-$  flow were similar in the presence and absence of  $\text{Na}^+$ . The diffusional flow of  $\text{HCO}_3^-$  was estimated after inhibition of  $\text{Na}^+$  transport by amiloride. Amiloride addition to the  $\text{Na}^+$ -containing Ringer's caused a large, rapid decrease in the short-circuit current while there was no change in short-circuit current in  $\text{Na}^+$ -free Ringer's after amiloride addition.

The exchange flows of  $\text{HCO}_3^-$  and  $\text{Cl}^-$  were calculated as the difference between the measured total and passive movements of the individual ions. The calculated rate of  $\text{Cl}^-$  exchange was  $0.23 \pm 0.06 \mu\text{mol/h}$  in  $\text{Na}^+$ -containing media and  $0.28 \pm 0.08 \mu\text{mol/h}$  in  $\text{Na}^+$ -free media. Calculated  $\text{HCO}_3^-$  exchange rates were  $0.25 \pm 0.12$  and  $0.31 \pm 0.05 \mu\text{mol/h}$  in  $\text{Na}^+$ -containing and  $\text{Na}^+$ -free Ringer's, respectively. The rate of  $\text{HCO}_3^-$  secretion was not statistically different from the rate of  $\text{Cl}^-$  absorption regardless of the  $\text{Na}^+$  concentration and the  $\text{Cl}^-/\text{HCO}_3^-$  exchange rate was the same in the absence of  $\text{Na}^+$  as in its presence.

A previous investigation with the turtle urinary bladder has shown that in the presence of  $\text{Na}^+$  in the bathing solutions,  $\text{Cl}^-$  transport occurred by a  $\text{Cl}^-/\text{HCO}_3^-$  exchange process and was not affected by ouabain [3].  $\text{HCO}_3^-$  secretion by the turtle bladder was also unaffected by amiloride [9]. The cortical collecting tubule of the rabbit is also capable of  $\text{HCO}_3^-$  secretion [10, 11]. Net  $\text{HCO}_3^-$  secretion by the collecting tubule is not altered by addition of ouabain and is increased after amiloride [11]. These results suggest that secretion of  $\text{HCO}_3^-$  by these tissues is independent of transepithelial  $\text{Na}^+$  transport. However, removal of  $\text{Na}^+$  from the solutions bathing the isolated collecting tubule abolishes net  $\text{HCO}_3^-$  secretion [11].

As similar experiments had not been previously attempted in turtle bladder, the present study was undertaken. In this study, as in a previous investigation from another laboratory [2],  $\text{Cl}^-$  absorption was not a function of  $\text{Na}^+$  concentration. The equality of  $\text{Cl}^-$  absorption and  $\text{HCO}_3^-$  secretion in the presence of  $\text{Na}^+$  also confirms previous results [3]. However, unlike the results in collecting tubule [11], we have demonstrated that the rate of  $\text{HCO}_3^-$  secretion is independent of  $\text{Na}^+$ . In addition, the rates of  $\text{Cl}^-$  absorption and  $\text{HCO}_3^-$  secretion by the turtle bladder are equal in  $\text{Na}^+$ -free media.

Two of the three modes of transepithelial  $\text{Cl}^-$  transport that have been shown to occur in epithelial tissues require  $\text{Na}^+$  [1].  $\text{Cl}^-$  transport across tissues such as frog skin [12] and toad bladder [13] occurs passively down the electrical gradient created by transepithelial  $\text{Na}^+$  transport. In a wide variety of epithelia [1],  $\text{Cl}^-$  movement against an electrochemical gradient is coupled to  $\text{Na}^+$  movement down its electrochemical gradient. The input of metabolic energy maintains the electrochemical gradient for  $\text{Na}^+$ , and  $\text{Cl}^-$  transport occurs as a 'secondary active' process [1]. Clearly,  $\text{Cl}^-$  transport across the turtle urinary bladder is not due to either of these processes. The results presented in this paper are most consistent with  $\text{Cl}^-$  transport across the urinary bladder of the turtle occurring via a  $\text{Cl}^-/\text{HCO}_3^-$  exchange system. The driving force for this system remains to be defined.

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## References

- 1 Frizzell, R.A. Field, M. and Schultz, S.G. (1979) *Am. J. Physiol.* 236, F1—F8
- 2 Gonzalez, C.F., Shamoo, Y.E. and Brodsky, W.A. (1967) *Am. J. Physiol.* 212, 641—650
- 3 Leslie, B.R., Schwartz, J.H. Steinmetz, P.R. (1973) *Am. J. Physiol.* 225, 610—617
- 4 Oliver, J.A., Himmelstein, S. and Steinmetz, P.R. (1975) *J. Clin. Invest.* 55, 1003—1008
- 5 Eyman, E.D. and Husted, R.F. (1978) *Proceedings of the Twenty-first Symposium on Circuits and Systems*, pp. 326—330, Western Periodicals Company, North Hollywood, CA
- 6 Steinmetz, P.R. and Lawson, L.R. (1971) *Am. J. Physiol.* 220, 1573—1580
- 7 Husted, R.F., Cohen, L.H. and Steinmetz, P.R. (1979) *J. Membrane Biol.* 47, 27—37
- 8 Brodsky, W.A., Durham, J. and Ehrenspeck, G. (1979) *J. Physiol.* 287, 559—573
- 9 Husted, R.F. and Steinmetz, P.R. (1979) *J. Pharmacol. Exp. Ther.* 210, 264—268
- 10 McKinney, T.D. and Burg, M.B. (1977) *J. Clin. Invest.* 60, 766—768
- 11 McKinney, T.D. and Burg, M.B. (1978) *J. Clin. Invest.* 61, 1421—1427
- 12 Ussing, H.H. (1960) *The Alkali Metal Ions in Biology*, Springer-Verlag, Berlin
- 13 Leaf, A. (1965) *Ergeb. Physiol. Biol. Chem. Exp. Pharmacol.* 56, 216—263